RIDA®QUICK IFX Monitoring

Art. No.: GN3041

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1. Intended use

For *in-vitro* diagnostics. This test is a lateral flow immunochromatographic assay for the quantitative detection of infliximab (IFX, Remicade®) and the biosimilars Remsima® and Inflectra® in human serum and plasma.

2. Summary and explanation of the test

Therapeutic Drug Monitoring

Infliximab (IFX) is a chimeric antibody that targets the pro-inflammatory cytokine TNF-alpha. The introduction of infliximab has revolutionized the treatment of chronic inflammatory diseases like inflammatory bowel disease (IBD), rheumatoid arthritis (RA) and spondyloarthritis. It has been shown that infliximab can induce deep remission and improve the patient's quality of life [1]. Some patients do not respond to IFX therapy upon induction (primary non-responders), while others lose response over time (secondary non-responders). [2]

A drug can only exert its pharmacologic effect when adequate concentrations are achieved in the circulation. The serum concentration of infliximab just before the next infusion, defined as the trough concentration, has been used for therapeutic drug monitoring (TDM). Recent data on TDM have shown that a good clinical response is associated with adequate trough concentrations in IBD [3] and RA [4, 5] patients. TDM may therefore be very instrumental to optimize treatment and to overcome secondary loss of response.

RIDA®QUICK IFX Monitoring uses a highly specific monoclonal antibody (MA-IFX6B7), which was isolated and characterized at the KU Leuven. It detects only infliximab (Remicade®) as well as the biosimilars Remsima® und Inflectra®.

3. Test principle

IFX is detected through the formation of an antibody-antigen-sandwich of MA-IFX6B7 and TNFα. This is made visible by the usage of marked colloidal gold nanoparticles. The generated signal is read out with the RIDA®QUICK SCAN II and the IFX concentration calculated by using the standard curve which is stored in the instrument.

4. Reagents provided

Each kit contains sufficient reagents for 25 tests.

Cassette	1 pc	25 test cassettes
Sample diluent	25 ml	Sample dilution buffer
Reagent | A	2.5 ml	Reagent A
Reagent | B	2.5 ml	Reagent B

4.1 Additionally available reagents

Controls for RIDA®QUICK IFX Monitoring can be ordered separately. RIDA®QUICK IFX Monitoring Control Set (Art. No. GP3041) contains 2 controls. They are used in the same way as samples and can be used to check the test reagents and test procedure.
Content of RIDA®QUICK IFX Monitoring Control Set

High control	1.2 ml	Batch specific, high positive control
Low control	1.2 ml	Batch specific, low positive control

5. Reagents and their storage

Store the kit at 2 - 8 °C. Kit contents are stable until the expiration date printed on product label. The reagents should only shortly be left at room temperature. After usage, they should directly be stored at 2 - 8°C. The quality of the product cannot be guaranteed after the expiration date. Likewise, the usability of the cassettes can no longer be guaranteed if the cassette packaging is damaged.

6. Additionally required reagents – required accessories

- Reaction tube
- Sample tube for sample suspension (two for each patient sample)
- Micropipettes with disposable tips 10 – 100 µl und 100 – 1000 µl
- Stopwatch
- Waste container with 0.5 % hypochlorite solution
- RIDA®QUICK SCAN II (available at R-Biopharm AG, Art. No.: ZRQS2-KD
- Vortex mixer

7. Precautions for users

For *in vitro* diagnostic use only.

This test must only be carried out by trained laboratory personnel. The guidelines for working in medical laboratories must be followed and the instructions for carrying out the test must be strictly adhered to.

Samples or reagents must not be pipetted by mouth and contact with injured skin or mucous membranes must be prevented. When handling the samples, wear disposable gloves and when the test is finished, wash your hands. Do not smoke, eat or drink in areas where samples or test reagents are being used.

The reagents contain NaN_3 as a preservative. This substance must not be allowed to come into contact with the skin or mucous membrane.

8. Sample collection and storage

In this assay, EDTA-plasma samples, citrate plasma samples and serum samples may be used. Following collection, the serum should be separated from the clot as quickly as possible to avoid hemolysis. Transfer the serum to a clean storage tube.

Samples can be stored at 2 - 8 °C for 3 - 4 days or at -20 °C for at least one year. Repeated freezing and thawing should be avoided.

9. Test procedure

9.1. General information

The samples, sample dilution buffer, reagents A and B, and the test strips must be brought to room temperature (20 - 25 °C) before use. Once used, the test strips may not be re-used. The test must not be carried out in direct sunlight. Excess reagents must not be returned to the vessels because this can result in contamination.

The RIDA®QUICK SCAN II must be switched on prior to the start of the test. The test method must be scanned on first use using the barcode reader and is then saved for further measurements using the RIDA®QUICK SCAN II.

The batch-specific parameters must also be scanned once for each batch prior to the start of the test.

The QR codes for the test method and for the batch-specific parameters can be found on the analysis certificate included with the kit (see also the RIDA®QUICK SCAN II Manual).

9.2. Preparing the sample tests

9.2.1. Diluting the sample

The measurement range of RIDA®QUICK IFX Monitoring is between 0.5 - 10 µg/ml with use of the standard dilution (maintenance therapy phase). The measurement range can be extended to 2 - 40 µg/ml through an additional dilution (induction phase).

a) Measuring the trough level concentration during the maintenance therapy phase

To measure the trough level concentration during the maintenance therapy phase (= starting with week 14 and following), dilute the sample to 1:50 and use this later in the test in a final dilution of 1:500.

Dilute 20 µl of the sample in 980 µl sample dilution buffer Sample diluent (1:50).
Then use 20 µl of the sample diluted to 1:50 in the test (see 9.2.2).

b) Measuring the trough level concentration during the induction therapy phase

To measure the trough level concentration during the induction therapy phase (= in week 0, 2, 4, and 6) or to measure concentrations > 10 µg/ml, dilute the sample to 1:200 and use this later in the test in a final dilution of 1:2,000.

Next dilute 20 µl of the sample in 980 µl sample dilution buffer Sample diluent (1:50).
The 1:50 dilution from the maintenance therapy phase can also be used for this step.
Next dilute 100 µl of this solution in 300 µl Sample diluent (1:4) so that overall the dilution of the initial sample is 1:200.
Then use 20 µl of the sample diluted to 1:200 in the test.

9.2.2 Incubating the sample

In a separate reaction vessel, mix 90 µl Reagent | A (blue liquid, bottle with blue lid) and 90 µl Reagent | B (yellow liquid, bottle with transparent lid). If multiple test strips are processed, the solution can also be used for several samples at the same time.

Mixture of Reagent | A (blue liquid) and Reagent | B (yellow liquid) will create a green-colored solution.

Pipette 20 µl of the diluted sample solution into the 180 µl of the mixture of reagent A and B, which is equivalent to a further dilution of the sample of 1:10 (see 9.2.1. a) and b)). In this way, the final dilution of the initial sample will be 1:500 in the maintenance therapy phase and 1:2,000 in the induction therapy phase. Mix the solutions thoroughly by inversion or vortexing to homogenize the sample mixture.

Next incubate the reaction mixture at room temperature for exactly **5 minutes**.

9.3. Sample testing

Remove the test cassette Cassette from the packaging and place it on a flat surface. 100 µl of the sample preparation from the reaction tube of step 9.2.2. are pipetted into the sample well of the test cassette.

The test result always has to be read after **15 minutes** via the RIDA®QUICK SCAN II. The time needs to be strictly adhered to.

Color development of the lines can change during the entire development time and after drying. The color of the lines can vary from red to blue-violet/gray as the strip dries.

Measurement before or after completion of the 15 minutes incubation time can lead to wrong results.

10. Quality control – Signs of reagent deterioration

The test can only be evaluated, if the test cassette is unharmed and there are no color changes or lines present before applying the sample suspension. The control line (marked with C on the test cassette) has to show up in every test run. In case this band is missing the following should be checked before repeating the test:

- Expiry date of the reagents and test cassette used

- Correct test procedure

- Contamination of reagents

If the control line is still not visible after repeating the test with a different test cassette contact the manufacturer or your local R-Biopharm distributor.

11. Evaluation and interpretation

The read out is performed on the RIDA®QUICK SCAN II (also see RIDA®QUICK SCAN II-manual).

Please note: If the sample has previously been diluted by a factor of 4 (final dilution 1:2000), the result of the RIDA®QUICK SCAN II must be multiplied by four in order to obtain the actual IFX concentration (in µg/g) in the blood.

The control line (marked with C on the test cassette) has to show up in each run. In case it is missing, the test has not been done correctly or the reagents were not ok (see paragraph 10).

The test line (marked with T on the test cassette) shows up depending on the infliximab concentration of the sample after different incubation times and with different intensities. Only after the total run time of 15 minutes the final test result can be determined by using the RIDA®QUICK SCAN II. The time point for the read out must be strictly adhered to. The bands can change during the total incubation time and may also change after drying. The color of the band can vary from red to blue-violet/grey.

12. Performance characteristics

12.1. Precision

12.1.1. Intra-assay precision

The intra-assay precision was tested using five references with 20 replications each. The IFX concentrations were determined using the RIDA®QUICK SCAN II, and the resulting mean values (MV), the standard deviations (SD), and the variation coefficients (VC) of the readings were calculated for each sample. The results are listed in the following table.

Reference	1	2	3	4	5
MV (µg/ml)	0.93	3.12	5.23	7.09	8.76
SD	0.10	0.51	0.82	0.74	0.89
VC (%)	11.2	16.5	15.6	10.4	10.2

12.1.2. Inter-assay precision

The inter-assay precision was tested using five references with 40 replications each. The tests were carried out by three different operators on ten different test days in two runs each day (morning and afternoon). The IFX concentrations were determined using the RIDA®QUICK SCAN II, and the resulting mean values (MV), the standard deviations (SD), and the variation coefficients (VC) of the readings were calculated for each sample. The results are listed in the following table.

Reference	1	2	3	4	5
MV (µg/ml)	0.95	2.77	4.52	6.28	8.31
SD	0.13	0.41	0.74	0.82	1.30
VC (%)	13.66	14.69	16.31	13.01	15.61

12.2. Analytical sensitivity

For the determination of the analytical sensitivity, three control samples were tested with one dilution series each in two different batches, and the IFX concentrations were determined using the RIDA®QUICK SCAN II.

The detection limit is less than 0.5 µg/ ml IFX.

12.3. Detection rate

12.3.1. Detection rate for Remicade®

Three samples were mixed with each of the four different Remicade® quantities, and the IFX concentrations were determined using the RIDA®QUICK SCAN II.
The mean detection rate is 100%. The results are listed in the following table.

Sample	(µg/ml)	Addition of IFX (µg/ml)	Measured value (µg/ml)	Target value (µg/ml)	Detection rate (%)
1	1.07	6.24	7.61	7.31	104
		1.56	2.47	2.63	94
		5.46	6.56	6.53	100
		3.90	5.32	4.97	107
Mean value					101
2	1.14	5.41	6.12	6.55	93
		4.64	5.88	5.78	102
		0.77	1.84	1.91	96
		3.86	5.30	5.00	106
Mean value					99
3	1.07	7.02	7.73	8.09	96
		2.34	3.45	3.41	101
		3.90	5.43	4.97	109
		3.12	4.16	4.19	99
Mean value					101

12.3.2. Detection rate for biosimilars

a) Detection rate for Remsima®

Three samples were mixed with each of the four different Remsima® quantities, and the IFX concentrations were determined using the RIDA®QUICK SCAN II. The mean detection rate is 106%. The results are listed in the following table.

Sample	(µg/ml)	Addition of IFX (µg/ml)	Measured value (µg/ml)	Target value (µg/ml)	Detection rate (%)
1	1.29	6.96	8.59	8.25	104
		1.74	2.93	3.03	97
		6.09	7.55	7.38	102
		4.35	6.18	5.64	110
Mean value					103
2	1.31	6.08	8.05	7.39	109
		5.21	6.89	6.52	106
		0.87	2.13	2.18	98
		4.34	6.27	5.65	111
Mean value					106
3	1.30	7.82	9.42	9.12	103
		2.61	4.21	3.91	108
		4.35	6.47	5.65	115
		3.48	5.20	4.78	109
Mean value					109

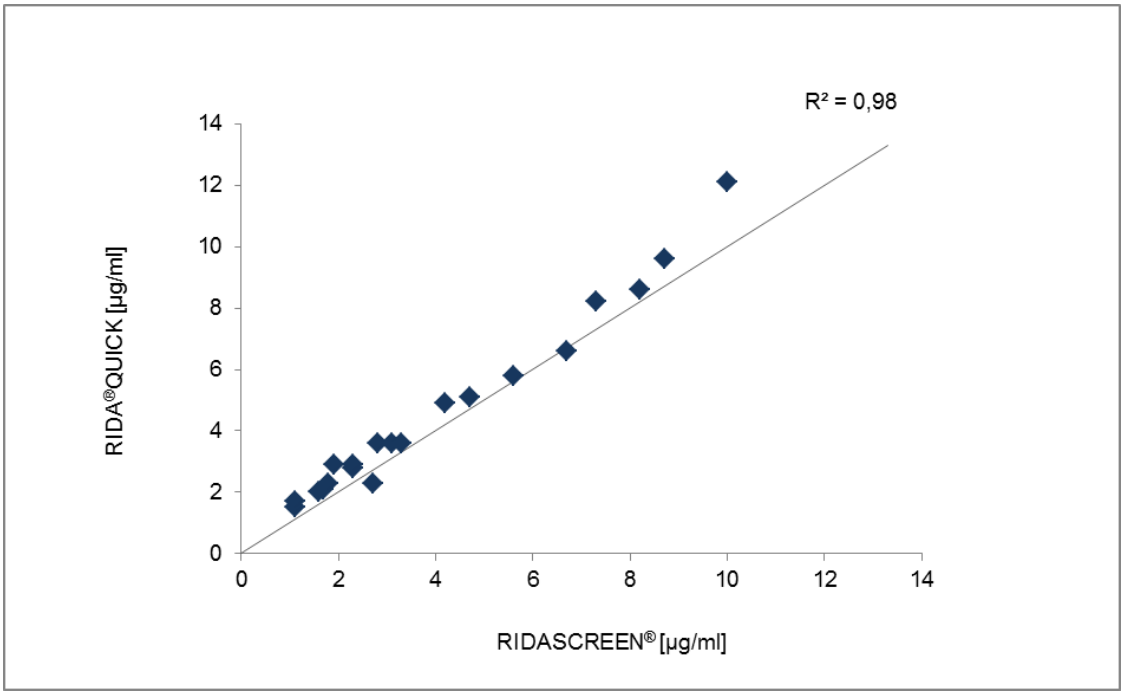
b) Detection rate for Inflectra®

Three samples were mixed with each of the four different Inflectra® quantities, and the IFX concentrations were determined using the RIDA®QUICK SCAN II. The mean detection rate is 98%. The results are listed in the following table.

Sample	(µg/ml)	Addition of IFX (µg/ml)	Measured value (µg/ml)	Target value (µg/ml)	Detection rate (%)
1	1.30	7.87	9.50	9.17	104
		1.97	3.22	3.27	98
		6.89	7.60	8.19	93
		4.92	6.14	6.22	99
Mean value					98
2	1.27	6.91	8.17	8.18	100
		5.92	7.18	7.19	100
		0.99	2.38	2.26	105
		4.94	6.45	6.21	104
Mean value					102
3	1.32	8.84	9.89	10.16	97
		2.95	4.03	4.27	94
		4.91	5.79	6.23	93
		3.93	4.94	5.25	94
Mean value					95

13. Correlation with reference assay

20 IFX-positive samples in the concentration range of 2 µg/ml to 12 µg/ml were measured in RIDASCREEN® IFX Monitoring and RIDA®QUICK IFX Monitoring, and the concentration was determined. The correlation coefficient was $R^2 = 0.98$. The results are shown in the following illustration.



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